

## Ecological habitat of small mammals as reservoirs of *Leishmania* spp. in the endemic foci of cutaneous leishmaniasis, the north Iran

Seyed Vahid Oranjin<sup>1</sup>, Haji Gholi Kami<sup>1\*</sup>, Farideh Tohidi<sup>2</sup>, Zeinolabedin Mohammadi<sup>3\*</sup>

<sup>1</sup> Department of Biology, Faculty of Sciences, Golestan University, Gorgan, Iran

<sup>2</sup> Laboratory Sciences Research Center, Golestan University of Medical Sciences, Gorgan, Iran

<sup>3</sup> Department of Biology, Farhangian University, Tehran, Iran

### Article type:

Original article

### Keywords:

Diversity

*Leishmania*

Iran

Rodents

Vector-borne

diseases

### Article history:

Received:

June 10, 2024

Revised:

August 23, 2024

Accepted:

August 24, 2024

Available online:

September 25, 2024

### Abstract

Small mammals are invasive species to agriculture and food resources worldwide. They are also reservoir hosts for medically and veterinary important zoonotic pathogens worldwide. Identifying ecological environments that serve as different small mammals' habitats as reservoirs for vector-borne diseases can help us to restrain their populations and manage the place of risk for spreading zoonotic diseases. This study was aimed to collect and identify the small mammals in northern Iran and to investigate the habitats with more potential risk for reservoirs of zoonotic *Leishmania*. In this survey, 294 specimens of small mammals were collected from the north of Iran between 2019 to 2023. Additionally, the abundance of zoonotic hosts and the rate of *Leishmania* prevalence reports in small mammals during the last ten years were analyzed to detect the relationship between reservoir distribution and parasitic infection. The collecting specimens belong to 12 families, 20 genera, and 26 species, and the most variety of species were collected from forest areas, while the lowest (4 species) were caught from urban and rural areas. According to the relative abundance index, the four most abundant small mammal species were *Rhombomys opimus*, *Rattus norvegicus*, *Mus musculus*, and *Microtus obscurus*. *Rhombomys opimus*, *R. norvegicus*, and *Meriones libycus* exhibit the highest levels of infection. This research, suggests comprehensive studies to detect *Leishmania* infection to determine new reservoirs in small mammals.

### Introduction

The north of Iran features a diverse range of ecological habitats like Hyrcanian forests, wetlands, and mountainous areas. Hyrcanian forests comprise a continuous 800-kilometers-belt of deciduous forests located between the Caspian Sea and the Alborz Mountains, with an area of over 1.8 million hectares (1). These forests were long known

as a biodiversity hotspot (Hyrcanian hotspot) and past refugia for several groups especially small mammals including orders Rodentia, Lagomorpha, Eulipotyphla, Chiroptera, and some species of Carnivora (2-5). Hyrcanian forests and the Alborz Mountains host a high diversity of small mammals (3, 5, 6). Small mammals such as rodents, bats, insectivores, hares, and some species of carnivores

\*Corresponding authors: [hgkami2000@yahoo.com](mailto:hgkami2000@yahoo.com) / [mohammadi.zeinal@gmail.com](mailto:mohammadi.zeinal@gmail.com)

<https://doi.org/10.22034/jzd.2024.18522>

[https://jzd.tabrizu.ac.ir/article\\_18522.html](https://jzd.tabrizu.ac.ir/article_18522.html)

Cite this article: Oranjin SV., Kami HGh., Tohidi F., and Mohammadi Z. Ecological habitat of small mammals as reservoirs of *Leishmania* spp. in the endemic foci of cutaneous leishmaniasis, the north Iran. Journal of Zoonotic Diseases, 2025, 9 (1): 717-729

Copyright© 2025, Published by the University of Tabriz.

This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY NC)



have been known as hosts and/or reservoirs of *Leishmania* spp. in the world. Iran is also an endemic country for leishmaniasis (7-10). Leishmaniasis is one of the vector-borne diseases caused by obligate intracellular parasites of the genus *Leishmania* (Kinetoplastida: Trypanosomatidae) that has been a severe health problem in tropical and subtropical areas including 98 endemic countries in the world and can infect over one billion people in those countries yearly (11). Leishmaniasis can be into Old World and New World Leishmaniasis. Old World Leishmaniasis can lead to Visceral Leishmaniasis (VL), caused by *Leishmania infantum* and *L. donovani*, and cutaneous leishmaniasis (CL), caused by *Leishmania major*, *L. tropica*, and *L. aethiopica* (12). This disease is transmitted by female sandflies (Diptera: Psychodidae: *Phlebotomus*) (13). Iran is one of the countries with the highest annual morbidity of Leishmaniasis in the world. On average, 20,000 new reports of CL are reported in Iran by the Ministry of Health per annum, and due to the high prevalence and endemicity of Cutaneous Leishmaniasis, the disease is one of the major community health problems (14, 15).

Additionally, rodents are known to remain asymptomatic carriers of *Leishmania* spp. for very long periods. Consequently, it can be hypothesized that rodent populations, as well as other wild animals, can maintain the permanent circulation of the parasite in an endemic area (16). The favorable ecological conditions and abundance of small mammals in the north of Iran, permit *Leishmania* species to spread into new geographical areas. Therefore, it is necessary to investigate possible species of reservoirs of *Leishmania* and their role in transmission.

Various species of small mammals such as *Rhombomys opimus*, *Nesokia indica*, *Meriones libycus*, *M. persicus*, *M. hurrianae*, *Tatera indica*, *Gerbillus nanus*, *Rattus rattus*, *R. norvegicus*, *Mus musculus*, and *Hemiechinus auritus* have been identified as potential reservoir hosts of the CL in Iran (8, 17, 18).

Some species of small mammals have been reported only from the north of Iran like *Crocidura capsica*, *Eptesicus nilssonii*, and *Martes martes* (3). This shows the diversity and uniqueness of the ecological habitats of northern Iran. The report of some species such as *R. opimus*, *M. libycus*, *N. indica*, *R. rattus*, *R. norvegicus*, and *M. musculus* as reservoirs of *Leishmania* and the presence of species of sand fly like *Phlebotomus papatasi* and *Phlebotomus sergenti* as *Leishmania* transmission agent, for instance in Golestan Province, in the northwest of Iran, *R. opimus sodalists* subspecies has been reported as a reservoir of Cutaneous Leishmaniasis (8) indicates the potential of other species of small mammals as a reservoir of *Leishmania* sp. in the north of Iran.

Distribution and community assemblage in small mammals have changed by anthropogenic activities in the area; therefore zoonotic diseases such as Leishmaniasis are dramatically increased. Recent studies show there is a significant correlation between the incidence of Cutaneous Leishmaniasis, the number of rodent burrows, the frequency of *Ph. papatasi* and geo-climatic factors (18, 19). For example, sand fly (*Ph. papatasi*) as the main vector of *L. major* has been collected from near rodent burrows such as *R. opimus*, *M. libycus*, *M. persicus*, and *Hemiechinus auritus* in Golestan Province, Iran (8, 17).

A variety of ecological habitats in the north of Iran supports a diverse population of mammalian species, making them potential hosts for *Leishmania* species. Therefore, studying their distribution and diversity is of critical importance. This study aimed to investigate the potential role of small mammals as reservoirs of *Leishmania* in northern Iran and elucidate the interrelations between wildlife biodiversity and dynamics of zoonotic disease in the north of Iran.

## Materials and methods

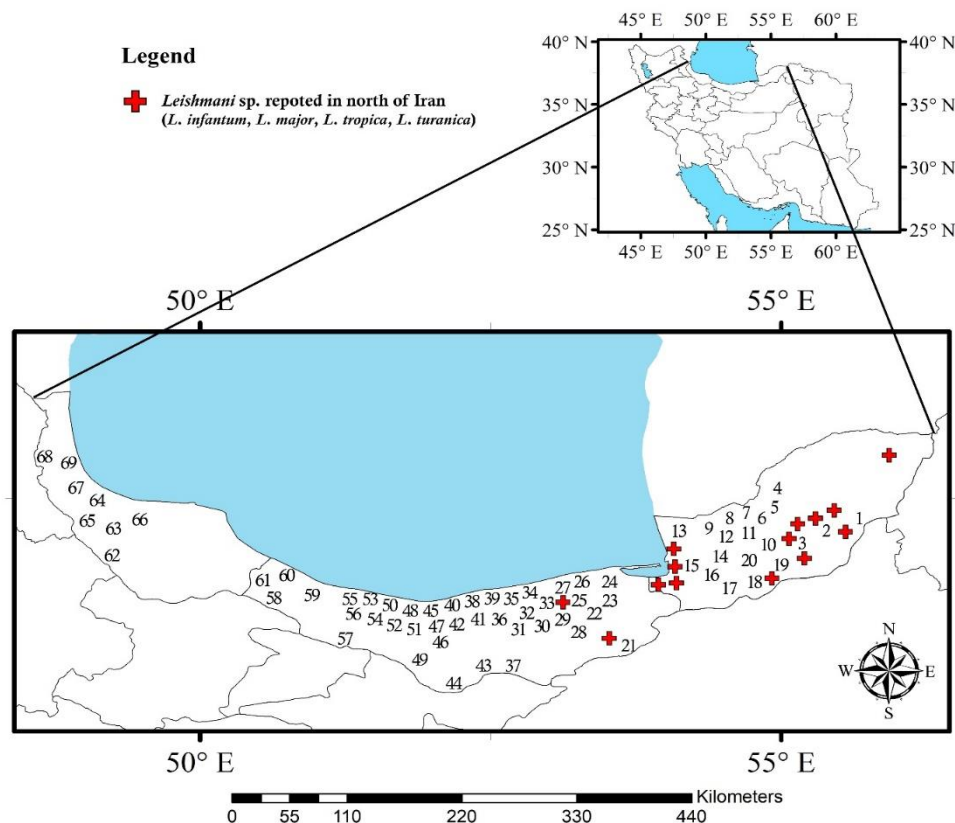
### Study area

The north of Iran includes different habitats, such as forests, mountains, and plains, which cause species

richness higher than other parts of Iran. This study was conducted in the eastern parts of the Hyrcanian region and the Elborz Mountains, spanning an elevation gradient from  $-20\text{m}$  (asl) to  $3000\text{m}$  (asl) from September 2019 to June 2023. The study area was partitioned into three elevation levels according to vegetation coverage: the first level ranged from  $-20$  to  $500$  m, encompassing farms and areas adjacent to wetlands; the second level spanned from  $500$  to  $1700$  m, covering deciduous forests of the Elborz Mountains range, mixed broadleaf forests, pasturage and coniferous scrublands; the third level stretched from  $1700$  to  $3000$  m, including the rocky areas of the Elborz Mountains. Sampling was conducted over 200 trap nights that included all three elevation levels.

#### Sampling

To investigate the present or absence of hosts as *Leishmania* reservoirs in northern Iran, 294 small mammals were captured using live traps and snack baits, pitfalls, and mist-net across five ecological habitats defined in this study including urban and rural areas, fields, and gardens, plain, forest, and mountainous areas (Table 1 and Figure 1). The snack, cucumber, and nuts were used as baits. The traps were placed at each station for a night and checked early in the morning. The captured specimens were collected in the morning and then were considered from sexed, weighed, and measured externally, and finally sacrificed specimens were fixed in ethanol 96%. The specimens were identified using diagnostic characters and identification keys available in the literature (20). ArcGIS ver.10.8 was used to prepare maps of the sampling area.



**Fig. 1.** Sampling map of small mammals and distribution of *Leishmania* in the small mammals and humans in the north of Iran. The specimens collected from 41 regions are marked by a number and reports of *Leishmania* in the small mammals and humans are marked by a plus on the map.

### Statistical analyses

The abundance of zoonotic hosts and the rate of Leishmaniasis prevalence were investigated to detect the relationship between host and parasite diversity. Host abundance was estimated as the total number of individuals captured per total capture as follows, in which the number captured by each species (total number of individuals captured) was divided by the total number of samples captured. We used the Web of Science to search for papers that reported Leishmaniasis in the north of Iran from 2013 to 2023 (Appendix). We examined experimental or observational studies between host quality and the presence of parasites in this area. Therefore, we used Analysis of Variance (ANOVA) to examine whether the host distributional range has a statistically significant effect on Leishmaniasis infection prevalence ( $P$ -value  $< 0.05$ ), also the abundance of each species was assessed using the relative abundance index. All statistical analyses were performed utilizing SPSS version 26.0 (SPSS, Chicago, IL, USA).

## Results

### Diversity and abundance of small mammals

In total, 294 specimens of small mammals belonging to 12 families, 20 genera, and 26 species were recorded and identified from the margins of the Caspian Sea in the north of Iran from September 2018 to June 2023 (Tables 1 and 2). The most prevalent species were *R. norvegicus*, *M. musculus*, and *M. obscurus*. According to the relative abundance index, *R. norvegicus*, representing 48 samples with an abundance of 16.43% among all samples, was found in four ecological habitats: plain areas, urban and rural areas, forest areas, and fields and gardens (Table 2 and Figure 2). *Microtus obscurus* and *M. musculus* were the second most abundant species with an abundance of 14.38% (42 specimens for each species). *Microtus obscurus* and *Calomyscus grandis* are new records for Gilan and Mazandaran Provinces respectively and were found in plain and mountainous areas (Table 2 and Figure 3), whereas *M. musculus* was found in forest and

urban areas. The species *A. hyrcanicus*, *C. grandis*, *H. indica* (the order Rodentia), and *P. kuhlii* (order Chiroptera) were rare in numbers, with an abundance of one specimen (0.34%) among all samples. *Apodemus hyrcanicus*, *H. indica*, and *P. kuhlii* were found in forest areas, while *C. grandis* was only observed in mountainous areas, with an elevation of over 2500 meters (Table 2 and Figure 3). *Calomyscus grandis*, *A. hyrcanicus*, and *Glis persicus* were endemic species of the research area. The highest number of species (16 species) were obtained from forest areas, while the lowest (4 species) were obtained from urban and rural areas (Figure 3). Therefore, the forest habitat type had the highest number of species and number of individuals, followed by the two ecological habitats of gardens and plants with 15 species.

### Records of small mammals as host of *Leishmania* spp. in the north Iran

From 2013 to 2023, 1056 samples of small mammals were captured and investigated from the north of Iran, of which 446 samples were infected. *Leishmania* infection was observed in Rodentia and Eulipotyphla specimens. *Rhombomys opimus*, *M. libycus*, *N. indica*, and *M. musculus* exhibit the highest levels of infection in north Iran.

The infection rates were investigated in the identified reservoirs in north Iran by species (CI = confidence interval: 95%; Table 3). These studies suggest that other small mammals, especially other rodents and eulipotyphla species, could also serve as potential *Leishmania* reservoirs in north Iran. The infection rates estimated in this survey were just significant for *M. persicus* (17.26-62.35%; CI = confidence interval: 95%; Table 3) meaning that the number of *M. persicus* sampled and the percentage of infection to the *Leishmania* parasite is not independence. The previous studies suggest that other small mammals, especially other rodent species e.g., could also serve as potential *Leishmania* reservoirs in northern Iran.

According to several studies from 2013 to 2023, four *Leishmania* species: *L. major*, *L. infantum*, *L. tropica*, and *L. turanica* have been reported in the

studied areas, the north Iran (Figure 1). *Leishmania tropica* and *L. major* species are dominant in Golestan Province, while, *L. infantum* and *L. major* species prevail in Mazandaran Province.

*Leishmania* infection has not been reported yet in human samples or possible reservoirs in Gilan (Guilan) Province and the west of Mazandaran until the time of compiling this research.

**Table 1.** Sampling locality, geographical coordinates, captured species, and their number at each region of the small mammals of the north of Iran, 2019–2023. Abbreviation: N: number; Lat: Latitude; Long: Longitude.

Province	Town	Sampling Locality	Num. of locality on the Fig. 1	Species	N	Lat.	Long.	
Golestan	HosseinAbad	Kalpush	1	<i>Myotis blythii</i>	5	37.18	55.75	
		Azad Shahr	Fazel Abad	2	<i>Lepus tibetanus</i>	2	37.06	55.20
		KhanBebin	Shirābād	3	<i>Rattus norvegicus</i>	3	36.98	54.99
	Inche Bouron				<i>Mus musculus</i>	1		
					<i>Mustela nivalis</i>	1		
			Tangeli	4	<i>Mus musculus</i>	3	37.45	54.71
			Ajigol	5	<i>Meriones libycus</i>	1	37.41	54.55
		Ashk Tappeh		6	<i>Meriones libycus</i>	3	37.43	54.60
					<i>Rhombomys opimus</i>	6		
		Aq Qala	Alagol Lake	7	<i>Rhombomys opimus</i>	22	37.42	54.62
			Inche Bouron	8	<i>Microtus sp.</i>	1	37.46	54.72
			Ghorban Abad	9	<i>Nesokia indica</i>	1	36.95	54.44
					<i>Mus musculus</i>	1		
	Agh Ghala		10	<i>Hemiechinus auritus</i>	1	37.04	54.46	
	Gorgan	Mohammad Abad		11	<i>Nesokia indica</i>	1	36.91	54.42
					<i>Rattus norvegicus</i>	1		
		Saqar Tappeh		12	<i>Mus musculus</i>	17	37.17	54.48
				<i>Microtus sp.</i>	8			
Torkaman Sahra		Mohammad Abad		13	<i>Lepus tibetanus</i>	7	37.09	54.19
				14	<i>Rattus norvegicus</i>	1	36.94	54.42
				<i>Mus musculus</i>	6			
		City Center	15	<i>Rattus norvegicus</i>	1	36.84	54.44	
		Marzan Kala	16	<i>Rattus norvegicus</i>	2	36.81	54.56	
		Gharn Abad	17	<i>Rattus norvegicus</i>	1	36.79	54.62	
Tuskestan				<i>Apodemus uralensis</i>	2			
				<i>Crocidura suaveolens</i>	8			
			18	<i>Rattus norvegicus</i>	7	36.79	54.57	
			<i>Apodemus uralensis</i>	1				
			<i>Apodemus hyrcanicus</i>	1				
	Shast Kola		19	<i>Mus musculus</i>	1	36.83	54.35	
				<i>Hemiechinus auritus</i>	1			
Mazandaran	Sari	Chale Bagh Park	20	<i>Pipistrellus kuhlii</i>	1	36.83	54.43	
		Kiasar	21	<i>Apodemus witherbyii</i>	1	36.17	53.71	
	Darab kola				<i>Ochotona rufescens</i>	1		
				22	<i>Crocidura sp.</i>	1	36.57	53.23
		Dashte Naz	23	<i>Erinaceus concolor</i>	1	36.82	53.17	
		Valuja	24	<i>Rattus norvegicus</i>	2	36.79	53.20	
		Chemaz Tappeh	25	<i>Mus musculus</i>	1	36.79	53.18	
		Shahrak Farhangian	26	<i>Rattus norvegicus</i>	1	36.48	52.94	
				<i>Mus musculus</i>	1			
	Qajar Kheyl		27	<i>Erinaceus concolor</i>	1	36.78	53.02	
		Amreh	28	<i>Pipistrellus pipistrellus</i>	2	36.42	53.11	
GhaemShahr	Siah Rood River			<i>Rhinolophus hipposideros</i>	1			
			29	<i>Rattus norvegicus</i>	1	36.45	52.88	

	Parchinak	30	<i>Rattus rattus</i>	1	36.45	53.01
	Eslam abad	31	<i>Rattus rattus</i>	1	36.46	52.88
			<i>Crocidura caspica</i>	1		
	Gharakheil	32	<i>Mus musculus</i>	4	36.46	52.77
	Vosta Kola	33	<i>Suncus etruscus</i>	5	36.37	52.50
			<i>Crocidura sp.</i>	6		
	Siahkola	34	<i>Apodemus uralensis</i>	1	36.44	52.85
	Ghadi Kola	35	<i>Glis persicus</i>	1	36.40	52.92
			<i>Crocidura sp.</i>	3		
	Berenjestanak	36	<i>Hystrix indica</i>	1	36.36	52.93
			<i>Erinaceus concolor</i>	1		
	Savadkooh	37	<i>Apodemus witherbyii</i>	2	35.96	52.67
Babol	Moqri Kala	38	<i>Rattus rattus</i>	1	36.37	52.60
	Babol Rood	39	<i>Rattus rattus</i>	1	36.37	52.72
			<i>Apodemus witherbyii</i>	1		
	Muziraj	40	<i>Mus musculus</i>	1	36.54	52.64
Amol	Dabudasht	41	<i>Rattus norvegicus</i>	1	36.48	52.45
	City Center	42	<i>Rattus norvegicus</i>	1	36.49	52.36
			<i>Apodemus witherbyii</i>	1		
	Larijan	43	<i>Apodemus witherbyii</i>	3	35.87	52.21
			<i>Calomyscus. grandis</i>	1		
	Malār	44	<i>Ochotona rufescens</i>	9	35.92	52.18
Mahmud Abad	Tileksar	45	<i>Mus musculus</i>	1	36.61	52.26
Chamestan	Kashpel park	46	<i>Rattus norvegicus</i>	1	36.48	52.11
			<i>Apodemus witherbyii</i>	2		
Nur	Shahrak Vahedi	47	<i>Rattus norvegicus</i>	3	36.55	51.99
	Louzan park	48	<i>Rattus norvegicus</i>	1	36.56	52.02
			<i>Apodemus witherbyii</i>	2		
	Baladeh	49	<i>Ochotona rufescens</i>	13	36.18	51.78
Royan	Shahrak Pasargad	50	<i>Rattus norvegicus</i>	1	36.53	51.99
			<i>Suncus etruscus</i>	2		
	Lale Royan Park	51	<i>Rattus norvegicus</i>	2	36.56	51.96
Chalandar	Molkar	52	<i>Mus musculus</i>	1	36.58	51.68
Shirud	Gil Mahale	53	<i>Apodemus witherbyii</i>	3	36.84	50.79
Noshahr	Shahr-e-Posht	54	<i>Rattus norvegicus</i>	3	36.62	51.50
	Musa Abad	55	<i>Suncus etruscus</i>	1	36.62	51.50
	Darzikola	56	<i>Erinaceus concolor</i>	1	36.59	51.52
Chaloos	Makarud	57	<i>Rattus norvegicus</i>	7	36.34	51.25
Tonekabon	2 Hezar	58	<i>Rattus norvegicus</i>	2	36.64	50.76
	Qaleh Gardan	59	<i>Rattus rattus</i>	3	36.75	50.83
	Lashgarak	60	<i>Apodemus witherbyii</i>	2	36.76	50.89
			<i>Crocidura caspica</i>	1		
Ramsar	Tosasan	61	<i>Rattus norvegicus</i>	2	36.92	50.64
Gilan	Masal	62	<i>Rattus norvegicus</i>	2	37.30	49.03
	Owlam	63	<i>Rattus norvegicus</i>	2	37.45	49.09
			<i>Rattus rattus,</i>	7		
			<i>Mus musculus,</i>	1		
			<i>Glis persicus</i>	2		
			<i>Rhinolophus</i>	3		
			<i>hipposideros</i>			
	Lowheh Sara	64	<i>Mus musculus</i>	2	37.36	49.13
	Duleh Malal	65	<i>Crocidura caspica</i>	6	37.36	49.15
	Shanbeh Rah	66	<i>Microtus obscurus</i>	33	37.29	48.93
	Sang Bast	67	<i>Crocidura caspica</i>	1	37.35	49.14
Navrud	Vardeh Sara	68	<i>Mustela nivalis</i>	1	37.69	48.98

**Table 2.** Ecological habitats of recorded and identified small mammals in the margins of the Caspian Sea, north of Iran.

Family	Species	Collection habitats				
		Plain areas	Urban and Rural areas	Forest areas	Mountainous areas	Fields and gardens
Muridae	<i>N. indica</i>	x	x			x
	<i>R. norvegicus</i>	x	x	x	x	x
	<i>R. rattus</i>	x	x	x		x
	<i>M. musculus</i>	x	x	x	x	x
	<i>A. witherbyii</i>	x		x	x	x
	<i>A. uralensis</i>			x		
	<i>A. hyrcanicus</i>			x		
	<i>M. libycus</i>	x				x
	<i>R. opimus</i>	x				x
Cricetidae	<i>M. obscurus</i>			x	x	x
Gliridae	<i>G. persicus</i>			x		x
Calomyscidae	<i>C. grandis</i>				x	
Hystriidae	<i>H. indica</i>			x		x
Soricidae	<i>S. etruscus</i>	x		x		x
	<i>C. caspica</i>	x		x		x
	<i>C. suaveolens</i>	x		x		x
	<i>Crocidura</i> sp.	x		x		x
Erinaceidae	<i>E. concolor</i>	x		x		x
	<i>H. auritus</i>	x				x
Leporidae	<i>L. tibetanus</i>	x				x
Ochotonidae	<i>O. rufescens</i>	x			x	
Vespertilionidae	<i>P. pipistrellus</i>			x		
	<i>P. kuhlii</i>		x	x		
	<i>M. blythii</i>			x		
Rhinolophidae	<i>R. hipposideros</i>			x		
Mustelidae	<i>M. nivalis</i>	x	x	x	x	x

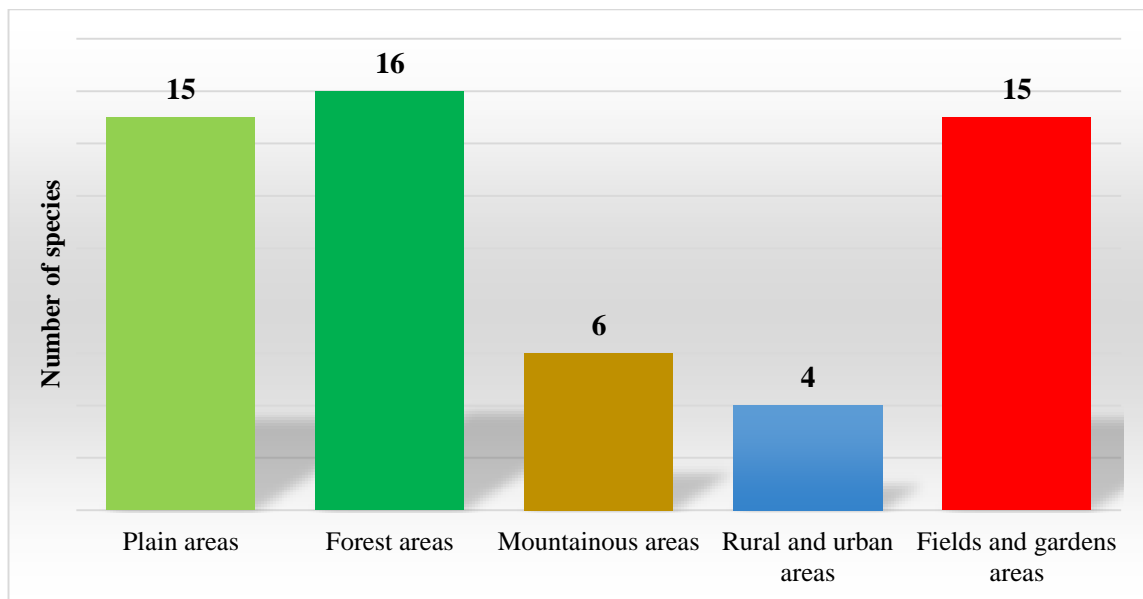


Fig. 2. Diagram of diversity of the captured species in the five ecological habitats.

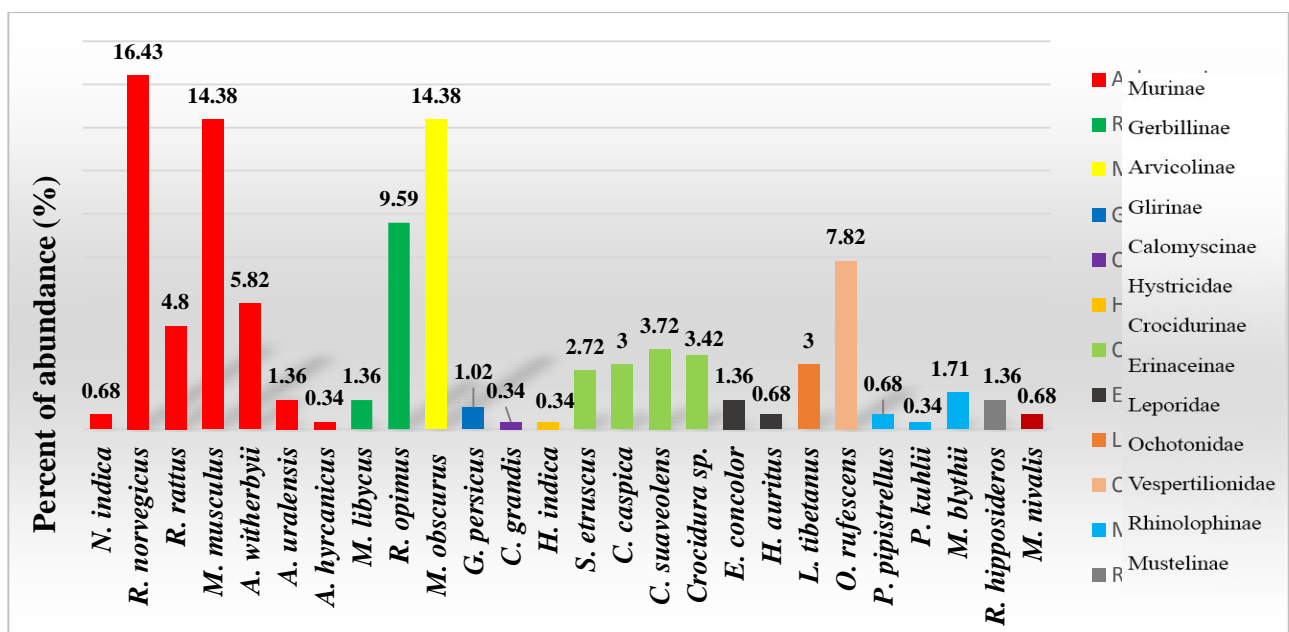


Fig. 3. Diagram of the abundance of captured samples per species (%), which are divided by different colors according to family and subfamily.

## Discussion

The survey of diversity, abundance, and distribution of terrestrial small mammals is vitally important to manage and control vector-borne diseases. Climate changes and anthropogenic activities are drivers for increasing vector-borne diseases worldwide.

Nowadays, there are concerns about altering vector host-pathogen relationships by these factors (21). There is an increased threat toward latitudes which are more affected by climate warming. Temperate regions are supposed to be the target of climate change effects on the relationship between biotic



factors causing diseases (22, 23). A rise in vector-borne diseases such as Leishmaniasis associated with small mammal reservoirs was documented in some studies. Carnivora, Rodentia, and Eulipotyphla (previously known as insectivores)

are the most small mammals being infected with *Leishmania* species (7, 9, 18, 24). Syanthropic rodents and the vicinity of rodent burrows in urban and rural areas can facilitate the transmission of CL to humans (25).

**Table 3.** The number of infection, percentage, and CI 95% infection species of the small mammals recognized as *Leishmania* sp. reservoirs in north Iran from 2013 to 2023. (T.N: Total number of investigated specimens for each species, N.I: Number of infected samples)

Species	T.N	N.I	Percent of infection	CI <sub>95%</sub>		P-Value
				Lower limit	Upper limit	
<i>Rhombomys opimus</i>	805	378	46.96%	40.56%	52.20%	0.021
<i>Meriones libicus</i>	134	50	37.31%	31.01%	41.12%	
<i>Mus musculus</i>	95	13	13.68%	10.68%	16.00%	
<i>Rattus norvegicus</i>	16	3	18.75%	11.70%	25.75%	
<i>Hemiechinus auratus</i>	6	2	33.33%	27.23%	38.98%	
<i>Rattus rattus</i>	-	-	-	-	-	
<i>Nesokia indica</i>	-	-	-	-	-	
<i>Meriones persicus</i>	-	-	-	-	-	
Total	1056	446	-	-	-	-

Since Iran is part of the geographical distribution of leishmaniasis many studies concentrated on the disease vectors and hosts in the different parts of the country. One systematic review and Meta-analysis reported Isfahan (66%) and Golestan (64%) Provinces as regions with the highest prevalence of CL associated with *L. major* while the lowest prevalence of CL was recorded from Kermanshah (4%), Hormozgan (10%), and Bushehr (12%) (26). In the spatial modeling of CL, from 1983 to 2013 represented by Holakouie-Naieni et al (27). Dry and desert climates in Iran used to be the endemic foci of the disease while the southwest of Iran was affected between 1983 and 1997, and subsequently, CL developed towards the center and eastern Iran between 1998 and 2013. Yazd, Khuzistan, and Kohgiluyeh-Boyer-Ahmad have also been supposed to be the hotspots for the disease. The results of both surveys and also present study are exactly fitted to the ecological niche modeling for the main reservoirs of CL (28). It is also noteworthy to mention that the occurrence of *Leishmania* in the north of Iran is mostly related to the presence of *R. opimus* and *N. indica* but in the central, south, and the western part of Iran, it is more associated with

the occurrence of the genus *Meriones*, *Tatera* and also *Rhombomys*. The amastigotes of *Leishmania* parasite have been also observed in *M. musculus* and *R. rattus* in south Iran (29, 30) and *M. persicus*, *T. indica*, and *M. musculus* were PCR-positive rodents for *Leishmania major* DNA (30, 31).

In general, the highest *Leishmania* infection rate has been reported from members of subfamilies Gerbilinae and Murinae as main reservoir hosts of Zoonotic Cutaneous Leishmaniasis (ZCL) in North Iran (7). Synanthropic rodents infected with *Leishmania* species were found in 20 percent of Iran, aslo *N. indica* and *R. opimus* show the highest prevalence with 48% and 39% infection, respectively (32). Rejali et al. in their meta-analysis demonstrated that Forest plots of CL prevalence (95% CI per 100,000 population) were 45.6 (44.1–47.2) (33). Moreover, there have been trends of increase in the incidence of CL in Iran from 2019 to 2022 (34). However, at the same time, it seems like the distribution of CL has been extended to a new area. In contrast, VL is mainly endemic in restricted regions of Iran, notably the northwest (Ardabil Province) and southwest (Fars Province) (35). Cutaneous Leishmaniasis is increasing in Iran

especially in the Golestan Province due to the extended distribution range of species from the subfamily Gerbilinae at endemic areas of ZCL (increasing animal reservoirs in a spatial and temporal) (36). For example, the distribution ranges of *R. opimus* and *M. libycus* have recently extended in the Caspian Sea lowland because of climatic changes and anthropogenic activities (37), which coincides with the increasing abundance of sand flies as well as *Leishmania* species in this area (26). As demonstrated in this study due to the presence of food and shelters, fields, gardens, and plain areas are much more occupied by potential reservoirs of *Leishmania*. In addition to climate change which may increase the length of suitable season for sandfly activities, the retreat of the Caspian Sea and left wetlands behind may increase the optimal area for sandfly spawning which was proposed to be more investigated. Moreover, deforestation and alteration of wild habitats have increased vector-host interaction and the risk of transmission of parasites to humans. Despite accessibility to food in the mentioned areas, lack of safety causes a rise in the population of rodents and other small mammals in urban and rural areas. On the other hand, the harsh climate of mountainous ranges and food shortages especially in cold seasons which are determinants for the living of small animals may cause low numbers of small mammals in mountains and their migration to urban and rural residential areas. Although bats have been reported as reservoirs of *Leishmania* throughout the world (38-40), there has not been a report of transmitting *Leishmania* into humans and livestock by bats in Iran yet. So the role of bats in the life cycle of *Leishmania* should be investigated in Iran. Natural infection of Hedgehogs to *L. major* has been previously reported (41). Hedgehogs have been also reported as reservoirs of CL in Iran. The result of the survey conducted by Yaghoobi-Ershadi et al. demonstrated that the rate of small mammals infection was high in *R. opimus* (82.1%), then *M. libycus* (15.7%), and last *H. auritis* (2.2%) in specimens collected from Isfahan (42).

The role of sand flies (Family: Phlebotominae) in the transmission of *Leishmania* is known (24). Several species of sand flies are distributed in the north of Iran some of these are known as the vectors of *Leishmania* like *Ph. papatasi*, and *Ph. Sergenti* (43-45). This issue can cause more transmission of *Leishmania* from the reservoirs to humans. According to the research done, it is suggested to identify new reservoirs, especially in the western areas of the northern strip of Iran. It is noteworthy to mention that stray dogs and other carnivores have been also reported as one of the main hosts for transmitting *Leishmania* parasites in vicinity to human.

Last but not least, during preparing the paper *Aedes* mosquitos and the viral infection of Dengue fever has been reported from different parts of Iran in 2024 (<https://www.who.int/>). *Aedes aegypti* and *Aedes albopictus*, the main vector for the Dengue virus may have also been coinfecting with *Leishmania* which has to be more investigated in Iran (46).

### Conclusion

Although there have been many studies on *Leishmania* reservoirs and vectors in Iran there were no comprehensive studies available to elucidate the role of each species of small mammals in spreading different kinds of *Leishmania* sp. Moreover, the importance of different small mammal ectoparasites as a vector of *Leishmania* is still vague which can be elucidated by applying a modern molecular approach. It is also proposed that the role of stray dogs in spreading leishmaniasis be studied. Additionally, a general survey to follow the effect of climate and ecological changes on the pattern of the leishmaniasis occurrence map is necessary. Mapping vector-reservoir ecology and disease incidence using upgraded modeling software is also recommended.

### Acknowledgments

We thank Dr. Fatemeh Ghorbani, Saleh Derkshan, and Najmeh Okhli for assistance in the field and laboratory works. We also thank Dr. Fatemeh Ghorbani for her comments on the initial version of

the manuscript. Permission to collect specimens was authorized by the Iranian Department of Environment (Permission Number: 99/420/16519).

### Ethical Approval

This study was conducted by the Declaration of Helsinki. Approval was granted by the ethical committee of Golestan University of Medical Sciences, Gorgan, Iran with ethics code, IR.GOUMS.REC.1399.072.

### Conflict of Interest

There is no conflict of interest.

### References

1. Talebi KS, Sajedi T, Pourhashemi M. Forests of Iran. A treasure from the past, a hope for the future. Illustrated, Netherlands. Springer Science & Business Media, 2013.
2. Naderi G, Kaboli M, Koren T, Karami M, Zupan S, Rezaei HR, et al. Mitochondrial evidence uncovers a refugium for the fat dormouse (*Glis glis* Linnaeus, 1766) in Hyrcanian forests of northern Iran. *Mamm Biol.* 2014; 79: 202-7. <https://doi.org/10.1016/j.mambio.2013.12.001>.
3. Zachos FE. The Atlas of Mammals of Iran. Mahmoud Karami, Taher Ghadirian and Kaveh Faizolahi. 2016. Iran Department of the Environment, Tehran. 292 pp. *J Animal Divers.* 2020; 2(2): 34-5. <http://doi.org/10.29252/JAD.2020.2.2.4>
4. Moradi A, Afsharzadeh S, Hamzehee B, Mozaffarian V. Study of plant diversity and floristics in the westernmost Hyrcanian forests. *J For Res.* 2020; 31: 1589-98. <http://doi.org/10.1007/s11676-019-00949-2>
5. Mohammadi Z, Ghorbani F, Aliabadian M, Lissovsky AA, Yazdani Moghaddam F, Olsson U. Multilocus phylogeny reveals habitat driven cryptic diversity in *Ochotona rufescens* (Ochotonidae). *Zool Scr.* 2022; 51(6): 617-28. <https://doi.org/10.1111/zsc.12564>.
6. Kafash A, Ashrafi S, Yousefi M. Biogeography of bats in Iran: Mapping and disentangling environmental and historical drivers of bat richness. *J Zool Syst Evol Res.* 2021; 59(7): 1546-56. <https://doi.org/10.1111/jzs.12520>.
7. Azami-Conesa I, Gómez-Muñoz MT, Martínez-Díaz RA. A Systematic Review (1990–2021) of Wild Animals Infected with Zoonotic Leishmania. *Microorganisms.* 2021; 9(5). <https://doi.org/10.3390/microorganisms9051101>.
8. Nasiri Z, Kalantari M, Mohammadi J, Daliri S, Mehrabani D, Azizi K. Cutaneous leishmaniasis in Iran: A review of epidemiological aspects, with emphasis on molecular findings. *Parasite.* 2022; 29: 47. <https://doi.org/10.1051/parasite/2022047>
9. Shahabi S, Azizi K, Asgari Q, Sarkari B. Calomyscid Rodents (Rodentia: Calomyscidae) as a Potential Reservoir of Zoonotic Cutaneous Leishmaniasis in a Mountainous Residential Area in the Plateau of Iran: Inferring from Molecular Data of kDNA and ITS2 Genes of *Leishmania Major*. *J Trop Med.* 2023; 2023: 5965340. <https://doi.org/10.1155/2023/5965340>.
10. de Souza NN, Ursine RL, Cruz DS, Xavier EMS, Queiroz LDRP, Falcão LAD, et al. *Leishmania* species infection of bats: A systematic review. *Acta Trop.* 2023; 248: 107025. <https://doi.org/10.1016/j.actatropica.2023.107025>.
11. Rai T, Shrestha S, Prajapati S, Bastola A, Parajuli N, Ghimire PG, et al. *Leishmania donovani* persistence and circulation causing cutaneous leishmaniasis in unusual-foci of Nepal. *Sci Rep.* 2023; 13(1): 12329. <https://doi.org/10.1038/s41598-023-37458-6>.
12. Rezaei Z, Pourabbas B, Asaei S, Sepehrpour S, Ahmadiania Motlagh S, Pourabbas P, et al. Livestock infected with *Leishmania* spp. in southern Iran. *Parasit Vectors.* 2022; 15(1): 215. <https://doi.org/10.1186/s13071-022-05313-8>.
13. Hooshyar H, Rasti S, Rostamkhani P. Cutaneous Leishmaniasis Lesion on the Ear from Kashan, Central Iran: A Case Report. *Iran J Parasitol.* 2023; 18(1): 119-124. <https://doi.org/10.18502/ijpa.v18i1.12390>.
14. Moradzadeh R, Golmohammadi P, Ashraf H, Nadrian H, Fakoorziba MR. Effectiveness of Paromomycin on Cutaneous Leishmaniasis in Iran: A Systematic Review and Meta-Analysis. *Iran J Med Sci.* 2019; 44(3): 185-95.
15. Khosravi A, Sharifi I, Fekri A, Kermanizadeh A, Bamorovat M, Mostafavi M, et al. Clinical Features of Anthroponotic Cutaneous

- Leishmaniasis in a Major Focus, Southeastern Iran, 1994-2014. *Iran J Parasitol.* 2017; 12(4): 544-53.
16. Alcover MM, Riera MC, Fisa R. Leishmaniosis in Rodents Caused by *Leishmania infantum*: A Review of Studies in the Mediterranean Area. *Front Vet Sci.* 2021; 8:702687. <https://doi.org/10.3389/fvets.2021.702687>.
  17. Rouhani S, Mirzaei A, Spotin A, Parvizi P. Novel identification of *Leishmania major* in *Hemichinus auritus* and molecular detection of this parasite in *Meriones libycus* from an important foci of zoonotic cutaneous leishmaniasis in Iran. *J Infect Public Health.* 2014; 7(3): 210-7. <https://doi.org/10.1016/j.jiph.2013.12.002>.
  18. Pourmohammadi B, Mohammadi-Azni S. Molecular Detection of *Leishmania major* in *Hemichinus auritus*: A Potential Reservoir of Zoonotic Cutaneous Leishmaniasis in Damghan, Iran. *J Arthropod Borne Dis.* 2019; 13(3): 334-43.
  19. Hosseini Farash BR, Mohebbali M, Kazemi B, Fata A, Hajjaran H, Akhoundi B, et al. Validation of a mixture of rK26 and rK39 antigens from Iranian strain of *Leishmania infantum* to detect anti-*Leishmania* antibodies in human and reservoir hosts. *Sci Rep.* 2022; 12(1): 10426. <https://doi.org/10.1038/s41598-022-14490-6>.
  20. Ghorbani F, Mohammadi Z, Darvish J, Aliabadian M. Gradient of rodent species diversity across altitudes in Hyrcanian region, northeast Iran. *Zool Ecol.* 2014; 24(3): 192-8. <http://doi.org/10.1080/21658005.2014.935611>
  21. Gray JS, Dautel H, Estrada-Peña A, Kahl O, Lindgren E. Effects of climate change on ticks and tick-borne diseases in Europe. *Interdiscip Perspect Infect Dis.* 2009;2009:593232. <http://doi.org/10.1155/2009/593232>.
  22. Ogden NH, Lindsay LR. Effects of Climate and Climate Change on Vectors and Vector-Borne Diseases: Ticks Are Different. *Trends Parasitol.* 2016; 32(8): 646-56. <http://doi.org/10.1016/j.pt.2016.04.015>.
  23. Ogden NH. Climate change and vector-borne diseases of public health significance. *FEMS Microbiol Lett.* 2017; 364(19). <http://doi.org/10.1093/femsle/fnx186>.
  24. Cecílio P, Cordeiro-da-Silva A, Oliveira F. Sand flies: Basic information on the vectors of leishmaniasis and their interactions with *Leishmania* parasites. *Commun Biol.* 2022; 5(1): 305. <http://doi.org/10.1038/s42003-022-03240-z>.
  25. Sofizadeh A, Vatandoost H, Rassi Y, Hanafi-Bojd AA, Rafizadeh S. Spatial Analyses of the Relation between Rodent's Active Burrows and Incidence of Zoonotic Cutaneous Leishmaniasis in Golestan Province, Northeastern of Iran. *J Arthropod Borne Dis.* 2016; 10(4): 569-76.
  26. Sabzevari S, Teshnizi SH, Shokri A, Bahrami F, Kouhestani F. Cutaneous leishmaniasis in Iran: A systematic review and meta-analysis. *Microb Pathog.* 2021; 152: 104721. <http://doi.org/10.1016/j.micpath.2020.104721>.
  27. Holakouie-Naieni K, Mostafavi E, Bolorani AD, Mohebbali M, Pakzad R. Spatial modeling of cutaneous leishmaniasis in Iran from 1983 to 2013. *Acta Trop.* 2017; 166: 67-73. <http://doi.org/10.1016/j.actatropica.2016.11.004>.
  28. Gholamrezaei M, Mohebbali M, Hanafi-Bojd AA, Sedaghat MM, Shirzadi MR. Ecological Niche Modeling of main reservoir hosts of zoonotic cutaneous leishmaniasis in Iran. *Acta Trop.* 2016; 160: 44-52. <http://doi.org/10.1016/j.actatropica.2016.04.014>.
  29. Davami MH, Motazedian MH, Kalantari M, Asgari Q, Mohammadpour I, Sotoodeh-Jahromi A, et al. Molecular Survey on Detection of *Leishmania* Infection in Rodent Reservoirs in Jahrom District, Southern Iran. *J Arthropod Borne Dis.* 2014; 8(2): 139-46.
  30. Akhoundi M, Mohebbali M, Asadi M, Mahmodi MR, Amraei K, Mirzaei A. Molecular characterization of *Leishmania* spp. in reservoir hosts in endemic foci of zoonotic cutaneous leishmaniasis in Iran. *Folia Parasitol (Praha).* 2013; 60(3): 218-24. <http://doi.org/10.14411/fp.2013.024>.
  31. Parhizkari M, Motazedian MH, Asqari Q, Mehrabani D. The PCR-based detection of *Leishmania major* in *Mus musculus* and other rodents caught in southern Iran: a guide to sample selection. *Ann Trop Med Parasitol.* 2011; 105(4): 319-23. <http://doi.org/10.1179/136485911X12987676649827>.

32. Foroutan M, Khademvatan S, Majidiani H, Khalkhali H, Hedayati-Rad F, Khashaveh S, et al. Prevalence of *Leishmania* species in rodents: A systematic review and meta-analysis in Iran. *Acta Trop.* 2017; 172: 164-72. <http://doi.org/10.1016/j.actatropica.2017.04.022>.
33. Rejali M, Dashtaki NM, Ebrahimi A, Heidari A, Maracy MR. Cutaneous Leishmaniasis Based on Climate Regions in Iran (1998-2021): A Systematic Review and Meta-Analysis. *Adv Biomed Res.* 2022; 11: 120. [http://doi.org/10.4103/abr.abr\\_90\\_21](http://doi.org/10.4103/abr.abr_90_21).
34. Sharifi I, Khosravi A, Aflatoonian MR, Salarkia E, Bamorovat M, Karamoozian A, et al. Cutaneous leishmaniasis situation analysis in the Islamic Republic of Iran in preparation for an elimination plan. *Front Public Health.* 2023; 11: 1091709. <http://doi.org/10.3389/fpubh.2023.1091709>.
35. Sharifi I, Aflatoonian MR, Parizi MHD, Hosseininasab A, Mostafavi M, Bamorovat M, et al. Visceral Leishmaniasis in Southeastern Iran: A Narrative Review. *Iran J Parasitol.* 2017; 12(1): 1-11.
36. Tohidi F, Barghae A. Cutaneous leishmaniasis parasite identification via PCR in the infected areas in Golestan province. *Knowledge Health.* 2011; 6(2): 26-31.
37. Hamidi K, Mohammadi S, Ghassemi-Khademi T. Ecological niche modeling of genetic lineages of the great gerbil, *Rhombomys opimus* (Rodentia: Gerbillinae). *PLoS One.* 2021; 16(9): e0257063. <http://doi.org/10.1371/journal.pone.0257063>.
38. Kassahun A, Sadlova J, Benda P, Kostalova T, Warburg A, Hailu A, et al. Natural infection of bats with *Leishmania* in Ethiopia. *Acta Trop.* 2015; 150: 166-70. <http://doi.org/10.1016/j.actatropica.2015.07.024>.
39. Spitzová T. Transmission and epidemiology of visceral leishmaniasis. 2016.
40. Sandoval E. Identification of *Leishmania* spp. and *T. cruzi* Parasites in Bats Captured in El Paso, Texas Region: Bats as a New Reservoir. The University of Texas at El Paso; 2021.
41. Tomás-Pérez M, Khaldi M, Riera C, Mozo-León D, Ribas A, Hide M, et al. First report of natural infection in hedgehogs with *Leishmania major*, a possible reservoir of zoonotic cutaneous leishmaniasis in Algeria. *Acta Trop.* 2014; 135: 44-9. <http://doi.org/10.1016/j.actatropica.2014.03.018>.
42. Yaghoobi-Ershadi MR, Javadian E. Epidemiological study of reservoir hosts in an endemic area of zoonotic cutaneous leishmaniasis in Iran. *Bull World Health Organ.* 1996; 74(6): 587-90.
43. Norouzi B, Hanafi-Bojd AA, Moin-Vaziri V, Noorollahi A, Azari-Hamidian S. An Inventory of the Sand Flies (Diptera: Psychodidae) of Rudbar County, a New Focus of Leishmaniasis in Northern Iran, with a Taxonomic Note on the Subgenus *Larrousius*. *J Arthropod Borne Dis.* 2020; 14(3): 302-16. <http://doi.org/10.18502/jad.v14i3.4564>.
44. Sharbatkhori M, Spotin A, Taherkhani H, Roshanghalb M, Parvizi P. Molecular variation in *Leishmania* parasites from sandflies species of a zoonotic cutaneous leishmaniasis in northeast of Iran. *J Vector Borne Dis.* 2014; 51(1): 16-21.
45. Rafizadeh S, Saraei M, Abaei MR, Oshaghi MA, Mohebbali M, Peymani A, et al. Molecular Detection of *Leishmania major* and *L. turanica* in *Phlebotomus papatasi* and First Natural Infection of *P. salehi* to *L. major* in North-East of Iran. *J Arthropod Borne Dis.* 2016; 10(2): 141-7.
46. Coelho WMD, Bresciani KDS, Coelho JdCA, dos ANJOS LA, Buzetti WAS. Are *Aedes aegypti* mosquitoes potential vectors for leishmaniasis?—Case report. *Braz J Vet Res Anim Sci.* 2017; 54(4): 416-9. <http://doi.org/10.11606/issn.1678-4456.bjvr>

