

Short communication

Prevalence and pathologic changes due to *Sarcocystis* species in naturally infected sheep in Urmia city, Iran

Farhad Farhangpazhouh¹, Mohammad Yakhchali^{1*}, AmirAbbas Farshid², Hadi Rezaei¹

1- Department of Pathobiology, Parasitology division, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran

2- Department of Pathobiology, Pathology division, Faculty of Veterinary Medicine, Urmia University, Urmia, Iran

* **Corresponding Author:** *m.yakhchali@urmia.ac.ir*

(Received 14 July 2020, Accepted 17 Sep 2020)

Summary

Sarcocystis species in animals and humans cause a zoonotic disease called Sarcocystosis. This study was aimed to investigate the morphology and pathology of Sarcocystis in naturally infected sheep. The carcasses of slaughtered sheep at Urmia slaughterhouse were inspected for evidence of infection with *Sarcocystis* macrocysts. Histopathological sections were prepared and stained routinely by Hematoxylin-Eosin (H&E) staining. A total of 1372 out of 4121 (33.3%) removed macrocysts were full of bradyzoites (54.3×223.66µm) ranged from 5-10 mm. Histologically, the reaction of the muscle tissue varied from degenerative to inflammatory around the macrocysts. The infected muscle demonstrated evidence of myocytolysis with infiltration of inflammatory cells in focal pockets, which were mainly comprised of lymphocytes and macrophages. The other findings were arterial wall hyperplasia, hyperplastic proliferation, and giant cell presence around the macrocysts. The results indicated that morphologic features of macrocysts of *Sarcocystis* in naturally infected sheep were informative and causing pathologic changes in muscle tissue.

Keywords: Sheep, Carcasses, Macroscopic *Sarcocystis*, Histopathology, Iran

Introduction

Sarcocystosis is one of the zoonotic diseases and coccidian parasitic infections, which causes macrocysts (Chena et al., 2010). The parasites are found in muscles of intermediate host and infect the gastrointestinal tract in carnivores as the definitive host. To date, 35

out of 93 identified species reported from 200 species of mammals (Dalimi et al., 2008). Four species of *Sarcocystis* have been reported in sheep, including *S. tenella*, *S. arieticans*, *S. gigantean*, *S. medusiformis* (Dubey, 2010). Humans can also be intermediate hosts for a variety of other

Sarcocystis species, i.e., *S. suis* and *S. hominis*. In Iran, *Sarcocystis* infection in sheep was reported for the first time in 1972 (Chena et al., 2010; Dalimi et al., 2010). There were many reports on *Sarcocystis* infection throughout the world and Iran (Oryan et al., 1996; Razmi and Rahbari, 2000; Shekarforoush and Alikhani, 2003; Bonyadian and Meshki, 2006; Dalimi et al., 2010).

The conventional method of distinguishing *Sarcocystis* species and combining these data with information on the life cycle are not suitable due to little morphological variation, high antigenic cross-reactivity, and time-consuming (Motamedi et al., 2010). However, laboratory studies on *Sarcocystis* infection like histopathology reported the first in the brain of sheep in Iran in 2006 (Bonyadian and Meshki, 2006). Thus, the present study was carried out to determine the macrocyst of *Sarcocystis* species using laboratory studies.

Materials and methods

The esophagus, diaphragm, and skeletal muscles in carcasses of sheep were thoroughly inspected for the presence of *Sarcocystis* macrocysts at Urmia industrial abattoir. The macrocysts were removed, excised from the tissues, and identified based on the morphologic and morphometric characteristics.

Twenty grams of pooled muscles were digested in 50 mL of acid pepsin and incubated for 30 min at 40°C. The digested specimen was filtered through a fine-meshed sieve into a tube, centrifuged at 2000g for 5 min, and the sediment suspended in 0.5 mL of distilled water. The suspension was then microscopically examined for the presence of *Sarcocystis* bradyzoites under the light microscope at 400× magnification (Arshad et al., 2007; Dalimi et al., 2008).

All infected tissue muscles with *Sarcocystis* macrocysts were fixed in 10% formalin and paraffin. The tissues were sectioned in 4-5 μm thick pieces and stained with Hematoxylin and Eosin (H&E) staining and examined under a light microscope at 400×-1000× magnification (Dubey et al., 2000).

Results

The macrocysts of *Sarcocystis* occur as elongated cylindrical bodies, and milky-white colored cysts embedded in the muscular tissues with length ranged from <5 mm to >10 mm (Fig. 1). The inspected macrocysts were fat (29.3%, 1372/4121) in striated muscles of that were large enough to discriminate by the naked eye. The macrocysts were fully packed with banana-shaped bradyzoites averaging $5.43 \times 22.36 \mu\text{m}$ (range: 3.16 to 7.38×18.77 to $27.69 \mu\text{m}$) (Fig. 1). Based on this finding,

identified macrocysts maybe belonging to *Sarcocystis gigantea* (Fig. 1a-c).

Histopathological analysis of macrocysts of *Sarcocystis* revealed degenerative, myocytolysis, and inflammatory reaction and fibroblastic proliferation along with

fibroblast-like and inflammatory cell infiltration, *i.e.*, lymphocytes and macrophages (Fig.2a-c). The vascular lesion included vascular proliferation and wall hyperplasia in the infected muscle (Fig.2-b).

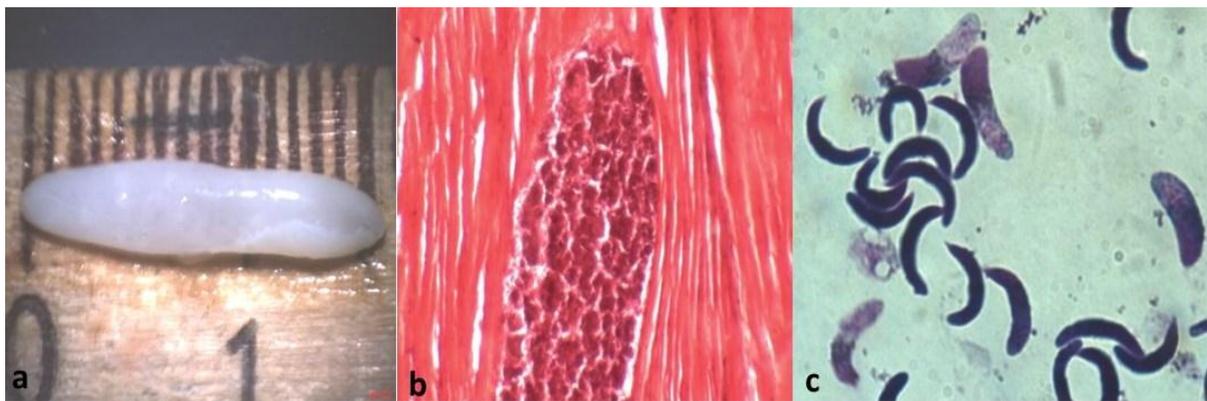


Fig. 1. A macrocyst of *Sarcocystis* in the diaphragm of naturally infected sheep (a), the cyst initiated no tissue reaction (b), bradyzoites (c) (H&E, 400×).

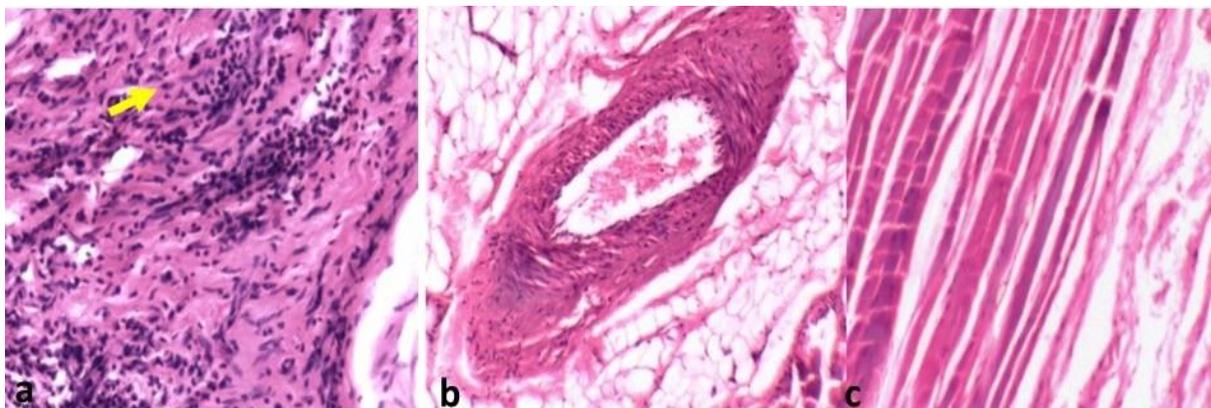


Fig. 2. Histologic sections of *Sarcocystis*: infected muscle showing evidence of myocytolysis (a), arterial wall hyperplasia in the infected muscle (b), part of a macrocyst along with inflammatory reaction, and fibroblastic proliferation (c), macrophage (yellow arrowhead) (H&E, 400×).

Discussion

Sarcocystis is a ubiquitous protozoan and causes a mild infection in sheep but is

important in meat inspection (Costa da Silva et al., 2009). In Iran, *S. gigantea* (synonyms: *S. ovifelis*) has spread worldwide, and its

oocysts are produced and excreted by cats and red foxes (Kojouri et al., 2011). Small ruminant infection with *S. gigantea* through close contact with cats and harboring oocysts in the pasture may be one of the main sources of high *Sarcocystis* infection in Iranian sheep (Dalimi et al., 2010). In most carcasses, infected animals with *Sarcocystis* macrocysts identified when doing meat inspection and observed between the muscle fibers (Yakhchali et al., 2010). In the present study, identified macrocysts as *S. gigantea* were in the form of elongated cylinders, which mainly reported from tissues of skeletal muscles of the diaphragm and esophagus. *Sarcocystis* macrocysts infection in sheep reports during the slaughterhouse inspections and laboratory studies (Razmi and Rahbari, 2000). During the meat inspection and chronic Sarcocystosis in sheep, *Sarcocystis* macrocysts were mostly present in the skeletal muscles of the esophagus and diaphragm (Oryan et al., 1996). In accordance with the current study, macroscopic lesion of *S. gigantea* in skeletal muscles was informing of strips of pale lines with petechia and ecchymosis. Fat atrophy around the heart and kidney, edema, focal necrosis in lymph nodes, and bleeding wounds in a serous visceral level also observed as described by Beyer (2001).

Microscopically, in the current study, the tissue muscles infected with *S. gigantea* were

in the form of inflammatory, degenerative, and myocytolysis around the macrocysts. There was no inflammatory cell infiltration, degenerative, and inflammatory lesions around the necrotic and degenerated microcysts in examined tissues. The influence of interstitial connective tissue caused the rupture of the muscle fibers. In this study, the focal accumulation of lymphocytes and macrophages, and vascular proliferation around the arterial vessel wall proposed. Dubey and Rommel (1992) noted that the lesions were microscopically in the form of hypertrophy of vascular endothelial cells, which were often along with infiltration of lymphocytes, neutrophils, macrophages, giant cells, few eosinophils, and plasma cells. In an earlier study, eosinophilic myositis due to *Sarcocystis* infection reported in cattle, sheep, pigs, and horses (Dubey et al., 1989). Valinezha et al. (2008) also reported infiltration of inflammatory cells like lymphocytes, macrophages, plasma cells, eosinophils, fibroblasts, and connective tissues around the degenerated cysts. One of the reasons for increasing in mononuclear cells around the vessels may probably be the host response to the antigen released from the sporozoite or immature schizont (Yarim et al., 2004; Radostits et al., 2007). The presence of antigens released by the parasite reported as a

reason for the infiltration of mononuclear cells (Oryan et al., 1996).

Conclusions

The present work demonstrated that laboratory studies on *Sarcocystis* macrocysts in sheep were informative and may found pathologic lesions in the skeletal muscle of sheep.

Acknowledgements

The authors wish to thank technical assistance of members in Pathobiology department, especially Mr. Badali and Mr. Morvaridi at Faculty of Veterinary Medicine, Urmia University.

Ethical approval

No applicable.

Conflict of interest statement

There is no conflict of interest.

References

- Arshad M., Dalimi A. and Ghaffarifar F. (2007). Comparative study on *Sarcocystis* diagnosis in meat of slaughtered sheep in Tabriz. *Pajouhesh & Sazandegi*, 75, pp. 68-72.
- Beyer T.V. (2001). Intracellular parasitism and the problem of *Sarcocystosis*. *Biology Bulletin*, 28(2), pp. 119-125.
- Bonyadian M. and Meshki B. (2006). Study on infestation of cow carcasses to *Sarcocyst* spp in slaughtered cows in Shahrekord by impression method. *Pajouhesh & Sazandegi*, 72, 14-18.
- Chena X., Zuoa Y., Rosenthalc B.M., Hed Y., Cuie L. and Yang Z. (2010). *Sarcocystis sinensis* is an ultrastructurally distinct parasite of water buffalo that can cause foodborne illness but cannot complete its life-cycle in human beings. *Veterinary Parasitology*, 178(1-2), pp.35-39.
- Costa da Silva R., Chunlei S.U. and Langoni, H. (2009). First identification of *Sarcocystis tenella* (Railliet, 1886), Moule 1886 (Protozoa: Apicomplexa) by PCR in naturally infected sheep from Brasil. *Veterinary Parasitology*, 165, pp.332-336.
- Dalimi A., Payekari H., Valizadeh M., Karimi Gh., Motamedi Gh., Abdi Ghodarzi M., Esmaeilzad M., Meshkat M. and Najar A. (2008). Detection of *Sarcocystis* spp. of slaughtered sheep in Gazvin Ziaran slaughterhouse by molecular assay. *Modares Journal of Medical Science*, 13(1 & 2), pp. 65-72.

- Dalimi A., Jalosian F., Tahvildar Biderouni F. and Ghaffarifar F. (2010). Identification of *Sarcocystis gigantea* by PCR- RFLP. *Journal of Veterinary Research*, 65(1), pp.43-46.
- Dubey JP. and Rommel M. (1992). Durch Protozoen bedingte Aborte bei landwirtschaftlichen Nutztieren. *DtschTierärztl Wochenschr*, 99, pp. 355-362.
- Dubey J. P., Saville W.J.A., Lindsay D.S., Stich R.W., Stanek J.F., Speer C. A., Rosenthal B.M., Njoku C.J., Kwok O.C.H., Shen S.K. and Reed S.M. (2000). Completion of the life cycle of *Sarcocystis neurona*. *Journal of Parasitology*, 86, pp.1276–1280.
- Dubey J.P. (2010). Two new species of *Sarcocystis* (Apicomplexa: sarcocystidae) Infecting the wolverine (*Gulo gulo*) from, Nunavut, Canada. *American Society Parasitology*, 96(5), pp. 972–976.
- Kojouri G.A., Aghajani E., Jahanabadi S. and Kojouri A. (2011). Mineral status of myocardial *Sarcocystosis*. *Iranian Journal of parasitology*, 6(2), pp.17-22.
- Motamedi Gh.R., Dalimi A., Aghaeipour K., Nouri A. (2010). Ultrastructural and molecular studies on fat and thin macrocysts of *Sarcocystis* spp. isolated from naturally infected goats. *Archives of Razi Institute*, 65 (2), pp. 91-97.
- Oryan A., Moghaddar N. and Gaur S.N. (1996). The distribution pattern of *Sarcocystis* species, their transmission and pathogenesis in sheep in Fars Province of Iran. *Veterinary Research Community*, 20(3), pp. 243-253.
- Radostits OM, Gay CC, Hinchcliff KW and Constable PD. (2007). *Veterinary medicine, A text book of the diseases of cattle, sheep, pig, goat, and horse*. 10th ed. Spain: Saunders, Elsevier.
- Razmi Gh. and Rahbari S. (2000). Survey on *Sarcocystosis* in domesticated ruminants slaughtered in Tehran and Golestan provinces. *Iranian Veterinary Journal*, 4, pp.39-46.
- Shekarforoush S.Sh. and Alikhani R. (2003). Prevalence of sarcocyst in slaughtered sheep in Isfahan, Iran. *Pajouhesh & Sazandegi*, 58, pp. 68-72.
- Valinezha A., Oryan A. and Ahmadi R. (2008). *Sarcocystis* and its complications in camels (*Camelus dromedarius*) of Eastern provinces of Iran. *Korean Journal of parasitology*, 46(4), pp. 229-234.

- Yakhchali M, Morshedi A. and Malekifar F. (2010). Screening of seropositive *Sarcocystis* (Apicomplexa: Sarcocystidae, Lankester 1882) in sheep using counterimmunoelectrophoresis. *Pajouhesh & Sazandegi*, 89, pp. 28-32.
- Yarim M., Yildiz K., Kabakci N. and Karahan S. (2004). Immunohistochemical localisation of 3 β -hydroxysteroid dehydrogenase in *Sarcocystis* spp.. *Parasitology Research*, 93, pp.457–460.
-