

Journal of Zoonotic Diseases

https://jzd.tabrizu.ac.ir/ Published by the University of Tabriz Online ISSN: 2717-2910



The report of *Mycobacterium avium subspecies paratuberculosis* in a wild goat (*Capra aegagrus*) in Iran

Mohammadreza Ghorani^{1*}, Fahime Eslami², Sina Soleimani³, Hossein Razi², Farbod Khakpour⁴

¹ Department of Pathobiology, Faculty of Veterinary Medicine, University of Tabriz, Tabriz, Iran

² Department of Environment, Chaharmahal and Bakhtiari Provincial Office, Shahrekord, Iran

in wildlife should be given special attention.

³ Razi Vaccine & Serum Research Institute, Agricultural Research, Education and Extension Organization (AREEO), Karaj, Iran

⁴ Department of Clinical Sciences, Faculty of Veterinary Medicine, Islamic Azad University-Shahrekord Branch, Shahrekord, Iran

Wild and domestic ruminants become infected with *Mycobacterium avium subsp.* paratuberculosis (MAP) leads to chronic enteritis, known as Johne's disease (JD). An

eight-year-old male wild goat (*Capra aegagrus*) that lived in the wildlife breeding center

in Chaharmahal and Bakhtiari province with symptoms of prolonged diarrhea that did not respond to antibiotic treatment was referred to a veterinary center. After a while, the

animal died. JD was diagnosed after laboratory diagnosis (by acid-fast staining). The

present study reported the incidence of JD in a wild goat in Iran for the first time. The

economic losses and animal health should be considered. Since 2020, the wild goat

(Capra aegagrus) has been classified in the category Near Threatened (NT) near treated

on the list (International Union for Conservation of Nature) IUCN. Therefore, protecting

the health of these species is essential. The importance of carrier animals and reservoirs

Article type:

Case report

Keywords:

Mycobacterium avium subsp. paratuberculosis Wild goat Johne's disease, Iran Article history: Received: May 9, 2023 Revised: November 6, 2023 Accepted: June 10, 2023 Available online: December 23, 2023

Introduction

Mycobacterium avium subsp. paratuberculosis (MAP) is a rod-shaped, small, aerobic, acid-fast, and intracellular bacterium of the Mycobacterium avium complex (1) that is the causative agent of

Abstract

Johne's disease (JD), which is relatively resistant to harsh environments. MAP is a small bacilli bacterium that causes chronic disease of the intestine, cecum, and mesenteric lymph nodes (2). In a wide variety of free and captive artiodactyls,

Cite this article: Ghorani M., Eslami F., Soleimani S., Razi H., and Khakpour F. The report of *Mycobacterium avium subspecies* paratuberculosis in a wild goat (*Capra aegagrus*) in Iran. Journal of Zoonotic Diseases, 2024, 8 (2): 509-514. Copyright© 2024, Published by the University of Tabriz.

This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International (CC BY NC)

^{*}Corresponding author: mo_gh66@yahoo.com

https://doi.org/10.22034/jzd.2023.16456

https://jzd.tabrizu.ac.ir/article_16456.html

clinical paratuberculosis has been observed (3, 4). However, non-ruminant MAP infections have also been reported, such as odd-toothed hawkers, rodents, lagomorphs, macropods, carnivores, birds, and inhuman primates (5, 6).

Animals are significantly susceptible to infection at an early age (7). JD exists in animals in two forms. In the multibacillary or lepromatous form, the cytoplasm of macrophages is stuffed with bacilli; there is chronically diffused granulomatous enteritis (8). These forms can only be distinguished histopathologically. A mention is useful if pathogens' intermittent or permanent excretion is also discussed (9). Only adult animals older than two years are usually shown to have clinical signs. Descriptions of paratuberculosis in wild species are usually incomplete and include sporadic case reports. The infection is progressive, chronic, and resistant to treatment. The clinical disease does not develop in most infected animals, but the bacteria may be excreted. Clinically, sick animals lose weight and, in some species, develop diarrhea and may die. During this disease, MAP is excreted in feces and milk, and bacteria are transmitted through blood and lymph vessels from infected animals to other internal organs. The infection spreads to both the male and female genitals. Although MAP is not considered a pathogen in humans, discussions are ongoing on the possibility of this mycobacterium playing an important role in public health (2).

An extended granulomatous and enteritis resistant to treatment with or without diarrhea, that leads to gradual weight loss despite ineffective food absorption, is known as classical clinical paratuberculosis (10). Severe diarrhea and intermandibular edema are common in cattle's late stages. Clinical signs are limited to chronic weight loss, disordered appearance, worsening of the condition, and lethargy in sheep and goats. Softer feces or diarrhea are rare and may only appear in the final stages (11).

The clinical signs of the disease are primarily undetectable, but when the clinical signs become apparent, the animal quickly becomes sicker and may die from the disease (4). The disease course provides useful diagnostic clues, failure to respond to treatment, and positive acid-fast lesions at necropsy are seen as clinical signs in individual animals (4).

In the case of periodic vaccination, the risks of transmission, the clinical incidence of the disease, and the rate of bacterial excretion are reduced. Vaccinated animals and tuberculosis surveillance programs interfere with serologic testing for MAP due to nonspecific responses to tuberculin skin tests, and implementation is limited (12). Its proper control can be easily achieved by vaccination (13). Also, for infection control, accurate diagnosis of infected subclinical animals is necessary. The purpose of the study was to report the occurrences of JD in a wild goat in Iran.

Materials and methods

Case history

Wild goat (*Capra aegagrus*) is a species of wild goat that lives in the bush, forests, and rocky areas from Turkey to Pakistan. In October 2021, A wild goat (male, eight years old, weighing approximately 70 kg) lived in the wildlife breeding center in Chaharmahal and Bakhtiari province (south-west of Iran) (Figure 1) with symptoms of prolonged diarrhea that did not respond to antibiotic treatment. A poor body condition was found in the clinical examination. The wild goat was sent to a veterinary center. The goat died shortly after being transported to the veterinary center.

Sample collection

Feces samples were taken fresh from the animal and were quickly sent to the microbiology laboratory with dry ice.

Microbiological investigation

Ziehl-Nelsen-stained feces and intestinal mucosa smears were studied microscopically. Clusters of organisms of small acid-fast bacilli were found (Figure 2a). A smear for coccidia was also done.



Fig. 1. The wild goat (Capra aegagrus) in the wildlife breeding center (Chaharmahal and Bakhtiari province, Iran).

Results

For some parts of the small intestine, particularly in the distal ileum, apparent gross pathology had been observed during postmortem examination. The acid-fast rod-shaped bacteria were detected by acid-fast staining (Figure 2b). No coccidia was found in the direct fecal smear.

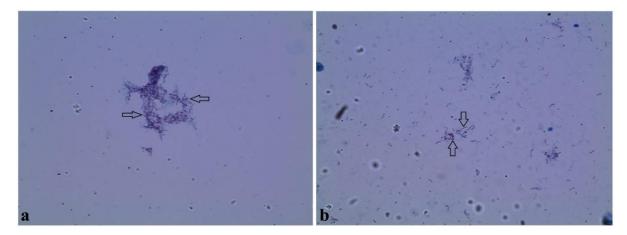


Fig. 2. a: Clusters of organisms of small acid-fast bacilli under the light microscope (×100 magnification). **b**: The acid-fast rod-shaped bacteria under the light microscope (×40 magnification).

Discussion

There is little information on the presence of MAP in different animal species in Iran. In this study, the presence of MAP was detected in a wild goat through necropsy and microscopic examinations. The tentative diagnosis was JD made based on clinical signs, supported later by the postmortem examinations. Although MAP species is not recognized as a zoonotic disease (14), the possibility of transmission of a common disease between humans and animals should not be ignored. Another significance of this report is that since 2020 the wild goat (*Capra aegagrus*) has been classified in the category NT near treated on the IUCN list. Therefore, protecting the health of these species is essential.

Because the information on JD in domestic animals is limited, therefore, it is not a priority disease to control in Iran, and the ongoing spread of JD among and within goat herds must be expected. Predictably, this will be the topic for future examinations. Few previous studies have reported the presence of MAP in wild ruminants. MAP occurrence in goats, sheep, cattle, buffaloes, and deer showed the ability to infect many animal species in the country. In Iran, Nassiri et al. identified 44% of fecal samples (from 243 specimens) and 18% of raw milk samples (from 56 specimens) of suspected cattle from some farms of Mashhad as infected with MAP (14). In another study, the MAP was detected in 15.1% of 212 fecal samples taken from four cattle farms in Kerman province, southeast of Iran (15).

Clinical cases with characteristic lesions on domestic dwarf goats of Western Africa have been mentioned in the disease reports from Germany's zoos (16). In a zoo in Missouri (USA), slow weight loss, thickening of animal hair, and diarrhea without odor were observed in Barbary sheep (Ammotragus lervia), and mouflon (Ovis orientalis) in adjacent areas with rapid disease progression and early death (17). After confirmation of fecal culture, necropsy, and histopathologic evaluation, a similar control regimen was administered in a top of Jimela (Damaliscus lunatus jimela), a subspecies of African antelope species in the USA, California (18). Dukes et al.'s report on zoos' problems with MAP described the spread of MAP in a herd of saiga antelopes (Saiga tatarica; Central Asian plains antelope) and their infection with mycobacterial infections in two zoos in Canada (Manitoba), has been explained (19). The risk of interspecific transmission between animals that share a home has also been reported from a zoo in

Turkey. In this report, the goats infected subclinically were identified by complement fixation and ELISA (20). A positive saiga antelope was reported by Orinbayev et al. In the Volga-Ural region of Kazakhstan, blood samples were collected from 286 free animals; the results showed that paratuberculosis in this species is also present in the natural population (21). MAP DNA has been detected in environmental samples as well as feces of snow goats (Oteroreamnos americanus) and pygmy goats at another zoo in Germany (22). In the study by Pourmahdi Borujeni et al., the seroprevalence of MAP in goats, sheep, and cattle in southwest Iran was compared. They announced no significant relationship between animal species and MAP infection (23).

Establishing an undeniable link and the risk of transmitting MAP from animals to humans should be considered an essential preventive measure. Therefore, developing vaccines and diagnostic systems to control MAP infection in the country's livestock population is essential.

Conclusion

Due to the risk of transmission of MAP from animals to humans, important preventive measures should be considered. Identifying the transmission of this pathogen from animals to humans allows for better evaluation of safety measures. These measures help to improve the level of public health. Therefore, developing vaccines and diagnostics systems to control MAP infection in the country's animal population is essential.

Acknowledgment

The authors thank the Chaharmahal and Bakhtiari Provincial Office of the Department of Environment.

Conflict of Interests

The authors declare that there is no conflict of interest.

Ethical approval

Not applicable.

References

- 1. Thorel MF, Krichevsky M, Lévy-Frébault VV. Numerical taxonomy of mycobactin-dependent emended description mycobacteria, of Mycobacterium avium, and description of Mycobacterium avium subsp. avium subsp. *Mvcobacterium* nov.. avium subsp. paratuberculosis subsp. nov.. and Mycobacterium avium subsp. silvaticum subsp. nov. Int J Syst Bacteriol. 1990 Jul; 40(3): 254-60. http://doi.org/10.1099/00207713-40-3-254
- 2. Ayele WY, Machackova M, Pavlik I. The transmission and impact of paratuberculosis infection in domestic and wild ruminants. Vet Med Czech. 2001; 46(7-8): 205-24. http://doi.org/10.17221/7878-VETMED
- 3. Weber R, Bryan RT, Schwartz DA, Owen RL. Human microsporidial infections. Clin Microbiol Rev. 1994 Oct; 7(4): 426-61. http://doi.org/10.1128/cmr.7.4.426
- 4. Roller M, Hansen S, Knauf-Witzens T, Oelemann WM, Czerny C-P, Abd El Wahed A, et al. *Mycobacterium avium* subspecies paratuberculosis infection in Zoo animals: A review of susceptibility and disease process. Front Vet Sci. 2020 Dec 23; 7: 572724. http://doi.org/10.3389/fvets.2020.572724
- Manning EJ. Paratuberculosis in captive and free-ranging wildlife. Vet Clin North Am Food Anim Pract. 2011 Nov; 27(3): 621-30. http://doi.org/10.1016/j.cvfa.2011.07.008
- Chittick E, Horne W, Wolfe B, Sladky K, Loomis M. Cardiopulmonary assessment of medetomidine, ketamine, and butorphanol anesthesia in captive Thomson's gazelles (Gazella thomsoni). J Zoo Wildl Med. 2001 Jun 32(2): 168-75. http://doi.org/10.1638/10427260
- Timms VJ, Gehringer MM, Mitchell HM, Daskalopoulos G, Neilan BA. How accurately can we detect *Mycobacterium avium* subsp. paratuberculosis infection? J Microbiol Methods.2011Apr;85(1):1-8. http://doi.org/10.1016/j.mimet.2011.01.026
- Debroy B, Tripathi B, Verma D. Pathology of paratuberculosis in sheep as confirmed by ISMav2 gene real-time polymerase chain reaction. Indian J Vet Pathol. 2010; 34(1): 17-22.
- 9. Catton BA. Paucibacillary paratuberculosis in a goat. Can Vet J. 2002 Oct; 43(10): 787-8.URL:

indianjournals.com/ijor.Aspx?target=ijor:ijvp &volume=34&issue=1&article=006

- Harris NB, Barletta RG. *Mycobacterium avium* subsp. paratuberculosis in veterinary medicine. Clin Microbiol Rev. 2001 Jul; 14(3): 489-512. http://doi.org/10.1128/CMR.14.3.489-512.2001
- Stehman SM. Paratuberculosis in small ruminants, deer, and South American camelids. Vet Clin North Am Food Anim Pract. 1996 Jul; 12(2): 441-55. http://doi.org/10.1016/s0749-0720(15)30416-3
- 12. Bastida F, Juste RA. Paratuberculosis control: a review with a focus on vaccination. J Immune Based Ther Vaccines. 2011 Oct 31; 9(1): 1-17. http://doi.org/10.1186/1476-8518-9-8
- Juste RA, Perez V. Control of paratuberculosis in sheep and goats. Vet Clin North Am Food Anim Pract. 2011 Mar; 27(1): 127-138. http://doi.org/10.1016/j.cvfa.2010.10.020
- Whiley H., Keegan A., Giglio S. & Bentham R. *Mycobacterium avium* complex–the role of potable water in disease transmission. J Appl Microbiol. 2012 Apr; 113(2): 223-232. http://doi.org/10.1111/j.1365-2672.2012.05298.x
- 15. Nassiri M, Jahandar MH, Soltani M, Mahdavi M, Doosti M. Identification and strain determination of M. paratuberculosis (MAP) by PCR and REA methods based on IS900 and IS1311 insertion segments. Agric Biotechnol J. 2012;4(1):83-96.

http://doi.org/10.22103/JAB.2012.469

- Soltani M. Detection of *Mycobacterium avium* subsp. paratuberculosis in Kerman Province's Dairy Cows using Microbial Culture, PCR and Nested PCR Methods. Iran J Animal Sci Res. 2018;10(2):263-73.
- http://doi.org/10.22067/IJASR.V10I2.65920 17. Seffner W, editor Paratuberkulose (Johne'sche Krankheit) bei Afrikanischen Zwergziegen. Erkrankungen der Zootiere: Verhandlungsbericht des VI Internationalen Symposiums über die Erkrankungen der Zoo-
- und Wildtiere Vienna; 1964. 18. Boever WJ. Johne's disease in aoudads and mouflon. J Zoo Animal Med. 1976; 7(1): 19-23. http://doi.org/10.2307/20094344
- 19. Steinberg H. Johne's disease (*Mycobacterium* paratuberculosis) in a Jimela topi (Damaliscus

lunatus jimela). J Zoo Animal Med. 1988; 33-41. http://doi.org/10.2307/20094850

20. Dukes TW, Glover GJ, Brooks BW, Duncan JR, Swendrowski M. Paratuberculosis in saiga antelope (Saiga tatarica) and experimental transmission to domestic sheep. J Wildl Dis. 1992Apr;28(2):161-70. http://doi.org/10/7580/0000/2558/28/2.161

http://doi.org/10.7589/0090-3558-28.2.161

- 21. Cihan H, Aytug N, Ozyigit M, Akcay E. Paratuberculosis in deer and small ruminants in a zoo in Turkey. Proc Eur Assoc Zoo Wildl Vet. 2006; 145-9.
- 22. Orynbayev MB, Beauvais W, Sansyzbay AR, Rystaeva RA, Sultankulova KT, Kerimbaev AA, et al. Seroprevalence of infectious diseases in saiga antelope (Saiga tatarica tatarica) in Kazakhstan 2012–2014. Prev Vet Med. 2016 May1;127:100-4. http://doi: 10.1016/j.prevetmed.2016.03.016. Epub 2016
- 23. Godin M GC, Fell S, Straubinger RK, editors. Vergleich zweier Methoden zur Detektion von Mycobacterium avium subsp. paratuberculosis DNA in Kot-und Umweltproben: 37 Arbeitstagung—Verband der Zootierärzte. VZT: Innsbruck; 2017
- 24. Pourmahdi Borujeni M, Haji Hajikolaei MR, Ghorbanpoor M, Elhaei Sahar H, Bagheri S, Roveyshedzadeh S. Comparison of *Mycobacterium avium subsp. paratuberculosis* infection in cattle, sheep and goats in the Khuzestan Province of Iran: Results of a preliminary survey. Vet Med Sci. 2021 Sep; 7(5):1970-1979.

http://doi.org/10.1002/vms3.559